

## DESCRIPTION

INHIBITOR OF LYMPHOCYTE ACTIVATION

## Technical Field

5           The present invention relates to a novel inhibitor of lymphocyte activation.

## Background Art

10           In the environment where humans live, there are numerous kinds of infectious microorganisms such as viruses, bacteria, fungi, parasites and the like. When growing in a living body each of these can cause a disease, eventually leading even to the death of the individual. Therefore, a defense mechanism for a living body against exogenous microorganisms is required in order for an individual to lead a healthy life. This mechanism is called "immunity."

15           The cells that are mainly responsible for immunity in the living body are lymphocytes. Lymphocytes are broadly classified into the T lymphocytes (T cells) and the B lymphocytes (B cells) by their functions. T cells are thought to have the ability of antigen presentation, cytotoxicity and the like, whereas B cells are believed to have the ability of antibody production. These two types of lymphocytes are derived from the same hemopoietic stem cells, which undergo various kinds of differentiation in the bone marrow or other organs or undergo repeated differentiation by the action of growth factors, and are finally released into the peripheral blood.

20           In the case of T cells, for example, hematopoietic stem cells differentiate into pre-T cells in the bone marrow and subsequently migrate to the thymus, where they undergo repeated differentiation to become mature T cells. Thereafter, they are activated by the stimuli of antigens to become activated T cells that have the ability of propagation, cytotoxicity and the like. In the case of B cells, on the other hand, hematopoietic

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stem cells differentiate via pro B cells and pre B cells into mature B cells by the stimuli of cytokines such as IL-1, IL-2, IL-4, IL-6 and the like in the bone marrow. They are then activated due to antigen challenge and finally become plasma cells that have an ability to produce antibodies.

Thus, the final activation is required for lymphocytes to develop their respective functions. As described above, because of its purpose of defending a living body against an exogenous foreign substance, immunity has a well-organized mechanism in which a foreign substance (the non-self) and the self are discriminatively recognized and a response is induced only against the non-self as antigen. However, when this mechanism is disrupted for some reason or other, even the self is recognized as antigen resulting in a disease called an autoimmune disease. When an immune response against the non-self takes place in an excessive or undesirable manner, tissues are impaired causing a state called allergy.

On the other hand, a reaction (rejection) that develops when an organ etc. of another person was transplanted to recognize and eliminate it as a non-self could be considered a normal biological mechanism. It is known that individuals are genetically different from each other, the representative of the difference being what is called the major histocompatibility complex (MHC). Transplantation of an organ from an individual having a different MHC could cause a severe rejection. Due to recent advances in the medical technologies, the necessity of transplantation of organs etc. has become very great in, for example, bone marrow transplantation in the treatment of leukemia or lymphoma, kidney transplantation in patients with terminal kidney diseases, transplantation of the cornea and the like. Concomitantly, the prevention of rejections has become a major challenge.

Due to extensive experiments and researches, pathways are being elucidated that lead from lymphocyte activation to a disease associated with it. Accordingly, many therapeutic drugs for diseases associated with lymphocyte activation have been developed. Among the autoimmune diseases caused by lymphocyte activation, currently rheumatoid arthritis, systemic lupus erythematosus, and scleroderma have been treated with non-steroidal anti-inflammatory drugs such as aspirin, steroids, or immunosuppressive agents such as azathioprine.

Furthermore, for the purpose of suppressing rejections associated with organ transplantation etc., immunosuppressive agents such as cyclosporine, azathioprine, and mizoribine have been used mainly in kidney transplantation and bone marrow transplantation. In addition, as therapeutic drugs for allergy, anti-histamine agents and inhibitors of the release of chemical transmitters that inhibit the liberation of chemical substances responsible for allergy have been used. Among them, however, none of the non-steroidal anti-inflammatory drugs, steroids, anti-histamine agents, and the inhibitors of the release of chemical transmitters act on the activation of lymphocytes responsible for allergy. They only represent symptomatic treatments of inflammation and thus do not essentially treat the diseases.

Due to their intrinsic properties, many of the immunosuppressive agents that are currently used have severe side-effects, such as decrease of blood corpuscles or shock, and hence cannot be considered adequate therapeutic agents. Moreover, even today there are no therapeutic regimens or drugs for many of the autoimmune diseases.

On the other hand, Goto, T. et al. have reported a monoclonal antibody (anti-HM1.24 antibody) that was obtained by immunizing mice with human myeloma cells

(Blood (1994) 84, 1922-1930). When anti-HM1.24 antibody was administered to a mouse transplanted with human myeloma cells, the antibody accumulated in tumor tissues in a specific manner (Masaaki Kosaka et al., Nippon Rinsho (Japan Clinical) (1995) 53, 627-635), suggesting that anti-HM1.24 antibody could be applied in the diagnosis of tumor localization by radioisotopic labeling, missile therapies such as radiotherapy, and the like. However, it was not known that anti-HM1.24 antibody is involved in the inhibition of lymphocyte activation.

#### Disclosure of the Invention

Therapeutic agents that are currently used for diseases associated with lymphocyte activation include various anti-inflammatory agents and immunosuppressive agents. As mentioned above, however, they are not completely satisfactory and therapeutic agents that can treat these diseases and alleviate the pains of the patient are being awaited. Thus, it is an object of the present invention to provide an inhibitor of lymphocyte activation.

In order to attain the above-mentioned purpose, the inventors have conducted intensive studies on anti-HM1.24 antibody (Goto, T. et al., Blood (1994) 84, 1922-1930) regarding its flow cytometry (FCM) analysis, its effects on blast formation by T cells, its effects on antibody production by B cells, and moreover on the isolation of the antigen protein to which anti-HM1.24 antibody specifically binds. As a result, the inventors have found that the antigen protein recognized by anti-HM1.24 antibody is expressed on the activated lymphocytes and that anti-HM1.24 antibody inhibits lymphocyte activation, and thereby have completed the present invention.

Thus, the present invention provides an inhibitor of lymphocyte activation comprising, as an active ingredient, an antibody that specifically binds to a protein having the amino acid sequence as set forth in

SEQ ID NO: 1.

The present invention also provides an inhibitor of T cell- or B cell-activation comprising, as an active ingredient, an antibody that specifically binds to a protein having the amino acid sequence as set forth in  
5 SEQ ID NO: 1.

The present invention also provides an inhibitor of lymphocyte activation comprising, as an active ingredient, a monoclonal antibody that specifically binds to a protein having the amino acid sequence as set forth  
10 in SEQ ID NO: 1.

The present invention also provides an inhibitor of lymphocyte activation comprising, as an active ingredient, an antibody that specifically binds to a protein having the amino acid sequence as set forth in  
15 SEQ ID NO: 1 and that has the constant region of human antibody.

The present invention also provides an inhibitor of lymphocyte activation comprising anti-HM1.24 antibody as an active ingredient.  
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The present invention also provides an inhibitor of lymphocyte activation comprising, as an active ingredient, a chimeric antibody or a humanized antibody.

The present invention also provides an inhibitor of lymphocyte activation comprising, as an active ingredient, a chimeric anti-HM1.24 antibody or a humanized anti-HM1.24 antibody.  
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The present invention also provides an inhibitor of lymphocyte activation comprising, as an active ingredient, an antibody that specifically binds to an epitope recognized by anti-HM1.24 antibody.  
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The present invention also provides a preventive and/or therapeutic agent for diseases associated with lymphocyte activation, comprising, as an active ingredient, an antibody that specifically binds to a protein having the amino acid sequence as set forth in  
35 SEQ ID NO: 1.

Furthermore, the present invention provides a preventive and/or therapeutic agent for autoimmune diseases, rejections in organ transplantation, and allergy, comprising, as an active ingredient, an antibody that specifically binds to a protein having the amino acid sequence as set forth in SEQ ID NO: 1.

#### Brief Description of the Drawings

Fig. 1 is a scheme showing that anti-HM1.24 antibody inhibits antibody production from B cells by SAC-stimulation.

Fig. 2 represents a histogram of FCM analysis of PHA-stimulated T cells with anti-HM1.24 antibody.

Fig. 3 represents a histogram of FCM analysis of activated T cells with anti-HM1.24 antibody.

Fig. 4 is a scheme showing that anti-HM1.24 antibody inhibits the blast formation reaction of PHA-stimulated T cells.

#### Embodiment for Carrying Out the Invention

##### 1. Antibody preparation

##### 1-1. Hybridoma preparation

Hybridomas that produce antibodies for use in the present invention can be basically constructed using a known procedure as described below. Thus, HM1.24 antigen protein or cells that express HM1.24 antigen may be used as sensitizing antigens and are used for immunization in the conventional method of immunization. The immune cells thus obtained are fused with known parent cells in the conventional cell fusion process, and then screened by the conventional screening method to select cells that produce monoclonal antibodies.

Specifically, monoclonal antibodies may be obtained in the following manner. For example, as a HM1.24 antigen-expressing cell which is a sensitizing antigen for obtaining antibody, there can be used a human multiple myeloma cell line KPMM2 (Japanese Unexamined Patent Publication (Kokai) No. 7-236475) or KPC-32 (Goto T. et al., Jpn. J. Clin. Hematol. (1991) 32, 1400).

Alternatively, as the sensitizing antigen, there may be used a protein having the amino acid sequence as set forth in SEQ ID NO: 1 or a peptide or polypeptide containing an epitope recognized by anti-HM1.24 antibody.

5           As used herein, cDNA that encodes a protein having the amino acid sequence as set forth in SEQ ID NO: 1 has been inserted in the XbaI cleavage site of pUC19 vector to construct plasmid pRS38-pUC19. E. coli having this plasmid has been internationally deposited under the provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pRS38-pUC19) on October 5, 1993 with the National Institute of Bioscience and Human Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-15 4434 (see Japanese Unexamined Patent Publication (Kokai) No. 7-196694). The cDNA fragment contained in this plasmid pRS38-pUC19 can be used to prepare a peptide or a polypeptide containing an epitope recognized by anti-HM1.24 antibody by a genetic engineering technology.

20           Preferably mammals to be immunized with the sensitizing antigen are selected in consideration of their compatibility with the parent cell for use in cell fusion. They generally include, but are not limited to, rodents such as mice, rats, hamsters and the like.

25           Immunization of animals with a sensitizing antigen is carried out using a known method. A general method, for example, involves the intraperitoneal or subcutaneous administration of a sensitizing antigen to a mammal. Specifically, a sensitizing antigen which has been 30 diluted and suspended in an appropriate amount of phosphate buffered saline (PBS) or physiological saline etc. is mixed, as desired, with an appropriate amount of Freund's complete adjuvant. After being emulsified, it is preferably administered to a mammal several times every 4 35 to 21 days. Alternatively a suitable carrier may be used at the time of immunization with the sensitizing antigen.

After immunization and the confirmation of the

increase in the desired antibody level in the serum, the immune cells are taken out from the mammal and are subjected to cell fusion, in which the preferred immune cells include, in particular, the spleen cells.

5       The mammalian myeloma cells as the other parent cells which are subjected to cell fusion with the above-mentioned immune cells preferably include various known cell lines such as P3X63Ag8.653) (J. Immunol. (1979) 123: 1548-1550), P3X63Ag8U.1 (Current Topics in Microbiology  
10       and Immunology (1978) 81: 1-7), NS-1 (Kohler, G. and Milstein, C., Eur. J. Immunol. (1976) 6: 511-519), MPC-11 (Margulies, D.H. et al., Cell (1976) 8: 405-415), SP2/0 (Shulman, M. et al., Nature (1978) 276: 269-270), FO (de St. Groth, S.F. et al., J. Immunol. Methods (1980) 35: 1-  
15       21), S194 (Trowbridge, I.S., J. Exp. Med. (1978) 148: 313-323), R210 (Galfre, G. et al., Nature (1979) 277: 131-133) and the like.

Cell fusion between the above immune cells and the myeloma cells may be essentially conducted in accordance  
20       with a known method such as is described in Milstein et al. (Kohler, G. and Milstein, C., Methods Enzymol. (1981) 73: 3-46) and the like.

More specifically, the above cell fusion is carried out in the conventional nutrient broth in the presence  
25       of, for example, a cell fusion accelerator. As the cell fusion accelerator, for example, polyethylene glycol (PEG), Sendai virus (HVJ) and the like may be used, and, in addition, an adjuvant such as dimethyl sulfoxide etc. may be added as desired to enhance efficiency of the  
30       fusion.

The preferred ratio of the immune cells and the myeloma cells to be used is, for example, 1 to 10 times more immune cells than the myeloma cells. Examples of culture media to be used for the above cell fusion  
35       include RPMI1640 medium and MEM culture medium suitable for the growth of the above myeloma cell lines, and the conventional culture medium used for this type of cell



culture, and besides a serum supplement such as fetal calf serum (FCS) may be added.

5 In cell fusion, predetermined amounts of the above immune cells and the myeloma cells are thoroughly mixed in the above culture medium, to which a PEG solution previously heated to about 37 °C, for example a PEG solution with a mean molecular weight of about 1000 to 6000, is added at a concentration of 30 to 60% (w/v) and mixed to obtain desired fusion cells (hybridomas). Then 10 by repeating the sequential addition of a suitable culture medium and centrifugation to remove the supernatant, cell fusion agents etc., which are undesirable for the growth of the hybridoma, can be removed.

15 Said hybridoma is selected by culturing in a conventional selection medium, for example, the HAT culture medium (a culture liquid containing hypoxanthine, aminopterin, and thymidine). Culturing in said HAT culture medium is continued generally for a period of 20 time sufficient to effect killing of the cells other than the desired hybridoma (non-fusion cells), generally several days to several weeks. The conventional limiting dilution method is conducted in which the hybridomas that produce the desired antibody are selected and monclonally 25 cloned.

In addition to obtaining the above hybridoma by immunizing an animal other than a human with an antigen, it is also possible to sensitize human lymphocytes in vitro with HM1.24 antigen or HM1.24 antigen-expressing 30 cells, and the resulting sensitized lymphocytes are fused with human myeloma cell, for example U266, to obtain the desired human antibody having the activity of binding to HM1.24 antigen or HM1.24 antigen-expressing cells (see Japanese Post-examined Patent Publication (Kokoku) No. 1- 35 59878). Furthermore, a transgenic animal having a repertoire of all human antibody genes is immunized with the antigen, i.e., HM1.24 antigen or HM1.24 antigen-

expressing cells, to obtain the desired humanized antibody, in the method described above (see International Patent Application WO 93/12227, WO 92/03918, WO 94/02602, WO 94/25585, WO 96/34096 and WO 96/33735).

The monoclonal antibody-producing hybridomas thus constructed can be subcultured in a conventional culture medium, or can be stored for a prolonged period of time in liquid nitrogen.

In order to obtain monoclonal antibody from said hybridoma, there can be mentioned a method in which said hybridoma is cultured in a conventional method and the antibodies are obtained in supernatant, or a method in which the hybridoma is administered to and grown in a mammal compatible with said hybridoma and the antibodies are obtained in the ascites. The former method is suitable for obtaining high-purity antibodies, whereas the latter is suitable for a large scale production of antibodies.

Specifically the anti-HM1.24 antibody-producing hybridoma can be constructed using: the method of Goto, T. et al. (Blood (1994) 84: 1922-1930). It can be conducted by a method in which the anti-HM1.24 antibody-producing hybridoma that was internationally deposited under the provisions of the Budapest Treaty as FERM BP-5233 on September 14, 1995 with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, is intraperitoneally injected to BALB/c mice (manufactured by CLEA Japan) to obtain the ascites from which the anti-HM1.24 antibody is purified, or: a method in which said hybridoma is cultured in a suitable culture medium such as the RPMI1640 medium containing 10% fetal bovine serum and 5% BM-Condimed H1 (manufactured by Boehringer Mannheim), the hybridoma SFM medium (manufactured by GIBCO-BRL), the PFHM-II medium (manufactured by GIBCO-BRL) and the like, and the anti-

HM1.24 antibody can be purified from the supernatant.

#### 1-2. Recombinant antibody

5 A recombinant antibody which was produced by the recombinant gene technology in which an antibody gene was cloned from the hybridoma and integrated into a suitable vector which was then introduced into a host can be used in the present invention as monoclonal antibody (see, for example, Carl, A.K., Borrebaeck, and James, W. Larrick, THERAPEUTIC MONOCLONAL ANTIBODIES, published in the  
10 United Kingdom by MACMILLAN PUBLISHERS LTD. 1990).

Specifically, mRNA encoding the variable region (V region) of the desired antibody is isolated from the hybridoma producing the antibody. The isolation of mRNA is conducted by preparing total RNA using, for example, a  
15 known method such as the guanidine ultracentrifuge method (Chirgwin, J.M. et al., Biochemistry (1979) 18, 5294-5299), the AGPC method (Chomczynski, P. et al., Analytical Biochemistry (1987) 162, 156-159), and then mRNA is purified from the total RNA using the mRNA  
20 Purification kit (manufactured by Pharmacia) and the like. Alternatively, mRNA can be directly prepared using the Quick Prep mRNA Purification Kit (manufactured by Pharmacia).

cdNA of the V region of antibody may be synthesized  
25 from the mRNA thus obtained using a reverse transcriptase. cdNA may be synthesized using the AMV Reverse Transcriptase First-strand cDNA Synthesis Kit and the like. Alternatively, for the synthesis and amplification of cdNA, the 5'-Ampli FINDER RACE Kit  
30 (manufactured by Clontech) and the 5'-RACE method (Frohman, M.A. et al., Proc. Natl. Acad. Sci. U.S.A. (1988) 85, 8998-9002; Belyavsky, A. et al., Nucleic Acids Res. (1989) 17, 2919-2932) that employs polymerase chain reaction (PCR) may be used. The desired DNA fragment is  
35 purified from the PCR product obtained and may be ligated to vector DNA. Moreover, a recombinant vector is constructed therefrom and then is introduced into E. coli

etc., from which colonies are selected to prepare a desired recombinant vector. The nucleotide sequence of the desired DNA may be confirmed by a known method such as the dideoxy method.

5        Once the DNA encoding the V region of the desired antibody has been obtained, it may be ligated to DNA encoding the constant region (C region) of the desired antibody, which is then integrated into an expression vector. Alternatively, the DNA encoding the V region of  
10       the antibody may be integrated into an expression vector which already contains DNA encoding the C region of the antibody.

         In order to produce the antibody for use in the present invention, the antibody gene is integrated as  
15       described below into an expression vector so as to be expressed under the control of the expression regulatory region, for example an enhancer and/or a promoter. Subsequently, the expression vector may be transformed into a host cell and the antibody can then be expressed  
20       therein.

### 1-3. Altered antibody

         In accordance with the present invention, artificially altered recombinant antibodies such as chimeric antibody and humanized antibody can be used for  
25       the purpose of lowering heterologous antigenicity against humans. These altered antibodies can be produced using known methods.

         Chimeric antibody can be obtained by ligating the thus obtained DNA encoding a V region of antibody to DNA  
30       encoding a C region of human antibody, which is then inserted into an expression vector and introduced into a host for production of the antibody therein (see European Patent Application EP 125023, and International Patent Application WO 96/02576). Using this known method,  
35       chimeric antibody useful for the present invention can be obtained.

         For example, E. coli having the plasmid that

contains DNA encoding an L chain V region or an H chain V region of chimeric anti-HM1.24 antibody has been internationally deposited under the provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pUC19-1.24L-gk) and Escherichia coli DH5 $\alpha$  (pUC19-1.24H-gyl),

respectively, on August 29, 1996 with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-5646 and FERM BP-5644, respectively (see Japanese Patent Application No. 9-271536).

Humanized antibody which is also called reshaped human antibody has been made by grafting the complementarity determining region (CDR) of an antibody of a mammal other than the human, for example mouse antibody, into the CDR of human antibody. The general recombinant DNA technology for preparation of such antibodies is also known (see European Patent Application EP 125023 and International Patent Application WO 96/02576).

Specifically, a DNA sequence which was designed to ligate the CDR of mouse antibody with the framework region (FR) of human antibody is synthesized by PCR method from several divided oligonucleotides having sections overlapping with one another at the ends thereof. The DNA thus obtained is ligated to the DNA encoding the C region of human antibody and then is integrated into an expression vector, which is then introduced into a host for antibody production (see European Patent Application EP 239400 and International Patent Application WO 96/02576).

FRs of human antibody linked through CDRs are selected so that the complementarity determining regions form a favorable antigen binding site. When desired, amino acids in the framework regions of the antibody variable region may be substituted so that the

complementarity determining region of reshaped human antibody may form an appropriate antigen binding site (Sato, K. et al., Cancer Res. (1993) 53, 851-856).

For example, E. coli having plasmid that contains a  
5 DNA encoding the version a (SEQ ID NO: 2) of the L chain V region and that for the version r (SEQ ID NO: 3) of the H chain V region of humanized anti-HM1.24 antibody has been internationally deposited under the provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pUC19-  
10 RVLa-AHM-gk) and Escherichia coli DH5 $\alpha$  (pUC19-RVHr-AHM-gyl), respectively, on August 29, 1996 with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-  
15 5645 and FERM BP-5643, respectively (Japanese Patent Application No. 9-271536). Furthermore, E. coli having plasmid containing a DNA encoding the version s (SEQ ID NO: 4) of the H chain V region of humanized anti-HM1.24 antibody has been internationally deposited under the  
20 provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pUC19-RVHs-AHM-gyl) on September 29, 1997 with the National Institute of Bioscience and Human Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as  
25 FERM BP-6127 (Japanese Patent Application No. 9-271536). For chimeric antibody or humanized antibody, the C region of human antibody is used, and most preferably human Cy can be used as the constant region of human antibody.

Chimeric antibody comprises the variable region of  
30 antibody derived from a mammal other than the human and the C region derived from human antibody, whereas humanized antibody comprises the complementarity determining regions of an antibody derived from a mammal other than the human and the framework regions (FRs) and  
35 the C region of antibody derived from human antibody.

Accordingly, antigenicity thereof in the human body has been reduced so that they are useful as the active ingredient of the therapeutic agents of the present invention.

5           A preferred embodiment of a humanized antibody for use in the present invention includes humanized anti-HM1.24 antibody (see Japanese Patent Application No. 9-271536). A preferred embodiment of an L chain V region of humanized anti-HM1.24 antibody includes one which has  
10           the amino acid sequence encoded by the nucleotide sequence as set forth in SEQ ID NO: 2. A preferred embodiment of the H chain V region of humanized anti-HM1.24 antibody includes one which has the amino acid sequence encoded by the base sequence as set forth in SEQ  
15           ID NO: 3 or 4.

#### 1-4. Expression and production

          Antibody genes constructed as described above may be expressed and the antibody can be obtained in a known method. In the case of mammalian cells, expression may be  
20           accomplished using an expression vector containing a commonly used useful promoter, an antibody gene to be expressed, and DNA in which the poly A signal has been operably linked at 3' downstream thereof or a vector containing said DNA. Examples of the promoter/enhancer  
25           include human cytomegalovirus immediate early promoter/enhancer.

          Additionally, as the promoter/enhancer which can be used for expression of antibody for use in the present invention, there can be used viral promoters/enhancers  
30           such as retrovirus, polyoma virus, adenovirus, and simian virus 40 (SV40), and promoters/enhancers derived from mammalian cells such as human elongation factor 1 $\alpha$  (HEF1 $\alpha$ ).

          For example, expression may be readily accomplished  
35           by the method of Mulligan et al. (Nature (1979) 277, 108) when SV40 promoter/enhancer is used, or by the method of

Mizushima et al. (Nucleic Acids Res. (1990) 18, 5322) when HEFl $\alpha$  promoter/enhancer is used.

5 In the case of E. coli, expression may be conducted by operably linking a commonly used useful promoter, a signal sequence for antibody secretion, and the antibody gene to be expressed, followed by expression thereof. As the promoter, for example, there can be mentioned lacZ promoter and araB promoter. The method of Ward et al. (Nature (1998) 341, 544-546; FASEB J. (1992) 6, 2422-10 2427) may be used when lacZ promoter is used, and the method of Better et al. (Science (1988) 240, 1041-1043) may be used when araB promoter is used.

15 As the signal sequence for antibody secretion, when produced in the periplasm of E. coli, the pelB signal sequence (Lei, S.P. et al., J. Bacteriol. (1987) 169, 4379) can be used. After separating the antibody produced in the periplasm, the structure of the antibody is appropriately refolded before use (see, for example, WO 96/30394).

20 As the origin of replication, there can be used those derived from SV40, polyoma virus, adenovirus, bovine papilloma virus (BPV) and the like. Furthermore, for the amplification of the gene copy number in the host cell system, expression vector can include as a selecttable marker an aminoglycoside transferase (APH) 25 gene, a thymidine kinase (TK) gene, an E. coli xanthine guaninephosphoribosyl transferase (Ecogpt) gene, the dihydrofolate reductase (dhfr) gene and the like.

30 For the production of antibody for use in the present invention, any production system can be used. The production system of antibody preparation comprises the in vitro or the in vivo production system. As the in vitro production system, there can be mentioned a production system which employs eukaryotic cells and the 35 production system which employs prokaryotic cells.

When the eukaryotic cells are used, there are the production systems which employ animal cells, plant



cells, and fungal cells. Known animal cells include (1) mammalian cells such as CHO cells, COS cells, myeloma cells, baby hamster kidney (BHK) cells, HeLa cells, and Vero cells, (2) amphibian cells such as *Xenopus* oocytes, or (3) insect cells such as sf9, sf21, and Tn5. Known plant cells include, for example, those derived from the genus Nicotiana, more specifically cells derived from Nicotiana tabacum, which is subjected to callus culture. Known fungal cells include yeasts such as the genus Saccharomyces, more specifically Saccharomyces cereviceae, or filamentous fungi such as the genus Aspergillus, more specifically Aspergillus niger.

When the prokaryotic cells are used, there are the production systems which employ bacterial cells. Known bacterial cells include Escherichia coli (E. coli), and Bacillus subtilis.

By introducing via transformation the gene of the desired antibody into these cells and culturing the transformed cells in vitro, the antibody can be obtained. Culturing is conducted in the known methods. For example, as the culture media, DMEM, MEM, RPMI1640, and IMDM can be used, and serum supplements such as fetal calf serum (FCS) may be used in combination. In addition, antibodies may be produced in vivo by implanting cells, into which the antibody gene has been introduced, into the abdominal cavity of an animal and the like.

Further, in vivo production systems, there can be mentioned those which employ animals and those which employ plants. When animals are used, there are the production systems which employ mammals and insects.

As mammals, goats, pigs, sheep, mice, and cattle can be used (Vicki Glaser, SPECTRUM Biotechnology Applications, 1993). Also, as insects, silkworms can be used.

When plants are used, tobacco, for example, can be used.

Antibody genes are introduced into these animals or

plants, and the antibodies are produced in such animals or plants, and recovered. For example, an antibody gene is inserted into the middle of the gene encoding protein which is inherently produced in the milk such as goat  $\beta$  casein to prepare fusion genes. DNA fragments containing the fusion gene into which the antibody gene has been inserted are injected into a goat embryo, and the embryo is introduced into a female goat. The desired antibody is obtained from the milk produced by the transgenic goat born to the goat who received the embryo or the offspring thereof.

In order to increase the amount of milk containing the desired antibody produced by the transgenic goat, hormones may be given to the transgenic goat as appropriate. (Ebert, K.M. et al., Bio/Technology (1994) 12, 699-702). When silkworms are used, baculovirus, into which a desired antibody gene has been inserted, is infected to the silkworm, and the desired antibody can be obtained from the body fluid of the silkworm (Susumu, M. et al., Nature (1985) 315, 592-594).

Moreover, when tobacco is used, a desired antibody gene is inserted into an expression vector for plants, for example pMON 530, and then the vector is introduced into a bacterium such as Agrobacterium tumefaciens. The bacterium is then infected to tobacco such as Nicotiana tabacum to obtain the desired antibody from the leaves of the tobacco (Julian, K.-C. Ma et al., Eur. J. Immunol. (1994) 24, 131-138).

When antibody is produced in vitro or in vivo production systems, as described above, DNA encoding the heavy chain (H chain) or the light chain (L chain) of antibody may be separately inserted into an expression vector and the hosts are transformed simultaneously, or DNA encoding the H chain and the L chain may be integrated into a single expression vector and the host is transformed therewith (see International Patent Application WO 94-11523).

The antibody produced as described above can be bound to various molecules such as polyethylene glycol (PEG) for use as a modified antibody. "Antibody" as used herein includes these modified antibodies. In order to obtain such a modified antibody, the antibody obtained may be chemically modified. These methods have already been established in the field of the art.

## 2. Separation of antibody and replication

### 2-1. Separation of antibody and replication

Antibodies produced and expressed as described above can be separated from the inside or outside of the cell or from the host and then may be purified to homogeneity. Separation and purification of the antibody for use in the present invention may be accomplished by affinity chromatography. As the column used for such affinity chromatography, there can be mentioned Protein A column and Protein G column. Examples of the carriers for Protein A column are Hyper D, POROS, Sepharose F.F. and the like.

Alternatively, methods for separation and purification conventionally used for proteins can be used without any limitation. Separation and purification of an antibody for use in the present invention may be accomplished by combining, as appropriate, chromatography other than the above-mentioned affinity chromatography, filtration, ultrafiltration, salting-out, dialysis and the like. Chromatography includes, for example, ion exchange chromatography, hydrophobic chromatography, gel-filtration and the like. These chromatographies can be applied into HPLC. Alternatively, reverse-phase chromatography can be used.

### 2-2. Determination of antibody concentration

The concentration of antibody obtained in the above 2-1 can be determined by the measurement of absorbance or by the enzyme-linked immunosorbent assay (ELISA) and the like. Thus, when absorbance measurement is employed, the antibody for use in the present invention or a sample

containing the antibody is appropriately diluted with PBS(-) and then the absorbance is measured at 280 nm, followed by calculation using the absorption coefficient of 1.35 OD at 1 mg/ml. When the ELISA method is used, measurement is conducted as follows. Thus, 100 µl of goat anti-human IgG (manufactured by BIO SOURCE) diluted to 1 µg/ml in 0.1 M bicarbonate buffer, pH 9.6, is added to a 96-well plate (manufactured by Nunc), and is incubated overnight at 4 °C to immobilize the antibody.

After blocking, 100 µl each of appropriately diluted antibody of the present invention or a sample containing the antibody, or 100 µl of human IgG of a known concentration as the standard is added, and incubated at room temperature for 1 hour. After washing, 100 µl of 5000-fold diluted alkaline phosphatase-labeled anti-human IgG antibody (manufactured by BIO SOURCE) is added, and incubated at room temperature for 1 hour. After washing, the substrate solution is added and incubated, followed by the measurement of absorbance at 405 nm using the MICROPLATE READER Model 3550 (manufactured by Bio-Rad) to calculate the concentration of the desired antibody.

### 3. Preparation of cells

The cells to be used in the present invention can be prepared according to the following methods.

#### 3-1. Preparation of human peripheral blood lymphocyte fraction

Peripheral blood, collected from healthy human donors and diluted 1/2 in PBS(-), is layered onto Ficoll-paque (manufactured by Pharmacia) in a 50 ml centrifuge tube (manufactured by Becton Dickinson). After centrifuge at 450 x g for 40 minutes at room temperature, the mononuclear cell fraction in the interface is isolated. After the fraction is prepared at a suitable density in a RPMI1640 medium (manufactured by GIBCO-BRL) containing 10% fetal bovine serum (manufactured by

Moregate), it is incubated under the condition of 37°C and 5% CO<sub>2</sub> for 1 hour in a plastic petri dish. The procedure is repeated twice to remove the cells attached to the dish. The remaining cells may be used in the following experiments as the human peripheral blood lymphocyte fraction.

3-2. Activation of human peripheral blood B cells by SAC

B cells in the peripheral blood lymphocytes can be activated by incubating the human peripheral blood lymphocytes prepared as above at a density of  $5 \times 10^6$  cells/ml with 0.01% SAC (Pansorbin cells, manufactured by Calbiochem.) in the presence or the absence of 1 ng/ml of IL-6 in a polypropylene tube under the condition of 37°C and 5% CO<sub>2</sub> for 2 days.

3-3. Purification of human peripheral blood T cells

Human peripheral T cells can be purified from the human peripheral blood lymphocytes prepared in the section 3-1 using the Collect Humm T cell Kit (manufactured by Biotex) according to the attached procedures.

3-4. Activation of human peripheral blood T cells by PHA

T cells in the peripheral blood lymphocytes can be activated by suspending the T cells prepared in the above section 3-3 in a RPMI1640 medium containing 2% fetal bovine serum (manufactured by Moregate) and then incubating the suspension at a density of  $1 \times 10^6$  cells/ml/well with the addition of 1 or 10 µg/ml PHA (Phytohemagglutinin, manufactured by Sigma) in a 24-well culture plate under the condition of 37°C and 5% CO<sub>2</sub> for 4 days.

4. FCM analysis

Reactivity of the antibody of the present invention with lymphocytes may be examined by flow cytometry (FCM) analysis. The cells used may be freshly isolated cells or the cultures thereof. As the freshly isolated cells,

there can be used, for example, peripheral blood mononuclear cells, peripheral blood lymphocytes, peripheral blood T cells, peripheral blood B cells and the like.

5           After washing the above cells in PBS(-), 100 µl of anti-HM1.24 antibody or a control antibody diluted to 25 µg/ml in the FACS buffer (PBS(-) containing 2% fetal bovine serum and 0.1% sodium azide) is added thereto, which is then incubated on ice for 30 minutes. After  
10       washing with the FACS buffer, 100 µl of 25 µg/ml FITC-labeled goat anti-mouse antibody (GAM, manufactured by Becton Dickinson) is added thereto, which is then incubated on ice for 30 minutes. After washing with the FACS buffer, the cells are suspended in 600 µl of the  
15       FACS buffer, and each cell may be measured for its fluorescence intensity using the FACScan (manufactured by Becton Dickinson).

#### 5. Confirmation of effects

          Lymphocyte activation is accompanied by blast  
20       formation in T cells and antibody production in B cells. Furthermore, with the activation of both cells, appearance or disappearance of various antigen markers on the cell surface is observed. Confirmation of the effects of inhibition of lymphocyte activation can be  
25       accomplished by inhibiting blast formation after adding the antibody for use in the present invention to the T cells, inhibiting antibody production after adding the  
- antibody for use in the present invention to the B cells, or by evaluating changes in the expression of antigen  
30       markers on the cell surface after adding the antibody for use in the present invention to the lymphocytes.

#### 5-1. Effects of anti-HM1.24 antibody on blast formation by T cells

          Effects of anti-HM1.24 antibody on blast formation  
35       by T cells can be evaluated by suspending the human peripheral T cells purified as described above in a

RPMI1640 medium containing 2% fetal bovine serum (manufactured by Moregate) and then incubating the suspension at a cell density of  $1 \times 10^5$  cells/200  $\mu$ l/well with 1  $\mu$ g/ml of PHA (Phytohemagglutinin, manufactured by Sigma) and 20  $\mu$ g/ml of anti-HM1.24 antibody or a control mouse IgG2a in a 96-well culture plate under the condition of 37°C and 5% CO<sub>2</sub> for 4 days. <sup>3</sup>H-thymidine (manufactured by Amersham) is added at 1  $\mu$ Ci/well and the incorporation of radioactivity after 4 hours may be measured using a  $\beta$ -counter (manufactured by Pharmacia).

5-2. Effects of anti-HM1.24 antibody on antibody production by B cells

Human peripheral blood lymphocytes prepared as described above are incubated at a density of  $5 \times 10^6$  cells/ml in a polypropylene tube with 0.01% SAC (Pansorbin cells, manufactured by Calbiochem.) under the condition of 37°C and 5% CO<sub>2</sub> for 2 days to activate the B cells in the peripheral lymphocytes. The SAC-treated peripheral blood lymphocytes are suspended in a RPMI1640 medium containing 10% fetal bovine serum (manufactured by Moregate), and the suspension at a cell density of  $1 \times 10^5$  cells/200  $\mu$ l/well is cultured with 20  $\mu$ g/ml of anti-HM1.24 antibody or a control mouse IgG2a in a 96-well culture plate (manufactured by Becton Dickinson) under the condition of 37°C and 5% CO<sub>2</sub> for 6 days, followed by collection of the culture supernatant.

The concentration of IgG in the culture supernatant can be measured by a human IgG-specific ELISA. Thus, 100  $\mu$ l of goat anti-human IgG (manufactured by TAGO) diluted to 1  $\mu$ g/ml in 0.1 M bicarbonate buffer (pH 9.6) is added to a 96-well immunoplate (manufactured by Nunc), and then incubated at 4 °C overnight to immobilize the antibody. After blocking, 100  $\mu$ l of appropriately diluted culture supernatant or human IgG (manufactured by CAPPEL) as a

standard is added thereto, and then incubated at room temperature for 1 hour.

After washing the plate, 100  $\mu$ l of 2,000-fold diluted alkaline phosphatase-labeled anti-human IgG (manufactured by CAPPEL) is added to the plate, and then incubated at room temperature for 1 hour. After washing the plate and adding the substrate solution thereto, it is incubated. Subsequently absorbance at 405 nm may be determined using the MICROPLATE READER Model 3550 (manufactured by Bio-Rad).

### 5-3. Analysis of antigen markers on the cell surface

Human peripheral blood lymphocytes or human peripheral T cells prepared as described above are cultured with PHA or SAC and anti-HM1.24 antibody or control mouse IgG2a as described in the section 5-1 or 5-2. These cells are reacted with antibodies capable of recognizing cell surface antigen markers that show changes in expression before and after activation such as CD10, CD25, CD38, CD40, CD47, CD54, CD98, PCA-1, HM1.24 antigens and the like. These can be subjected to FCM analysis as described in the above section 4.

### 5-4. Confirmation of effects and related diseases

It was revealed, as shown in the examples that follow, that the HM1.24 antigen is being expressed on the activated lymphocytes and that the addition of anti-HM1.24 antibody inhibited blast formation by T lymphocytes and furthermore antibody production by B lymphocytes. These facts indicated that anti-HM1.24 antibody has the effects of inhibiting the activation of lymphocytes.

On the other hand, as the diseases in which lymphocyte activation is involved, there can be mentioned autoimmune diseases, rejections associated with organ transplantation, and allergy. Specifically, autoimmune diseases include, for example, Hashimoto thyroiditis, primary myxedema, thyrotoxicosis, pernicious anemia, autoimmune atrophic gastritis, Addison disease, premature



menopause, insulin-dependent diabetes mellitus,  
Goodpature syndrome, myasthenia gravis, male infertility,  
pemphigus vulgaris, pemphigus, ophthalmia sympathica,  
lens induced uveitis, multiple sclerosis, autoimmune  
5 hemolytic anemia, idiopathic thrombocytopenic purpura,  
primary biliary cirrhosis, active chronic hepatitis,  
idiopathic cirrhosis, ulcerative colitis, Sjögren's  
syndrome, rheumatoid arthritis, dermatomyositis,  
scleroderma, mixed connective tissue diseases, discoid  
10 erythematosus, systemic lupus erythematosus and the like  
(translation supervised by Shunnichi Hirose et al.,  
Rinsho Mennekigaku Illustrated (Clinical Immunology  
Illustrated) (1994), Nankodo).

As rejections associated with organ transplantation,  
15 there are mentioned rejections associated with the  
transplantation of kidney, liver, and heart, epithelial  
or endothelial rejections associated with cornea  
transplantation, HVD, GVHD or the like associated with  
bone marrow transplantation (translation supervised by  
20 Shunnichi Hirose et al., Rinsho Men-ekigaku Illustrated  
(Clinical Immunology Illustrated) (1994), Nannkodo).  
Allergy includes, for example, type I allergy represented  
by atopic diseases, type II allergy observed in drug-  
related allergies, type III allergy that causes various  
25 nephritises, and type IV allergy represented by  
dermatitis caused by cosmetics or metals (Takeo Azeyanagi  
et al., Shin-Men-ekigaku Sosho (7) Men-eki to Allergy  
(New Immunology Series 7, Immunology and Allergy (1981),  
Igaku Shoin). Accordingly, the therapeutic agents of the  
30 present invention are useful as agents for treating  
diseases in which lymphocyte activation is involved.

#### 6. Route of administration and pharmaceutical preparation

The inhibitors of lymphocyte activation of the  
35 present invention may be administered, either  
systemically or locally, by a parenteral route, for  
example intravenous injection such as drip infusion,

intramuscular injection, intraperitoneal injection, and subcutaneous injection. The method of administration may be chosen, as appropriate, depending on the age and the condition of the patient. The effective dosage is chosen from the range of 0.01 mg to 100 mg per kg of body weight per administration. Alternatively, the dosage in the range of 1 to 1000 mg, preferably 5 to 50 mg per patient may be chosen. The inhibitors of lymphocyte activation of the present invention may contain pharmaceutically acceptable carriers or additives depending on the route of administration.

Examples of such carriers or additives include water, a pharmaceutical acceptable organic solvent, collagen, polyvinyl alcohol, polyvinylpyrrolidone, a carboxyvinyl polymer, carboxymethylcellulose sodium, polyacrylic sodium, sodium alginate, water-soluble dextran, carboxymethyl starch sodium, pectin, methyl cellulose, ethyl cellulose, xanthan gum, gum Arabic, casein, gelatin, agar, diglycerin, propylene glycol, polyethylene glycol, Vaseline, paraffin, stearyl alcohol, stearic acid, human serum albumin (HSA), mannitol, sorbitol, lactose, a pharmaceutically acceptable surfactant and the like. Additives used are chosen from, but not limited to, the above or combinations thereof depending on the dosage form.

#### Examples

The present invention will now be explained hereinbelow in more detail with reference to the following examples. It is to be noted that the present invention is not limited to these examples in any way.

#### Example 1. Construction of anti-HM1.24 antibody

##### 1. Preparation of mouse ascites containing anti-HM1.24 antibody

Hybridomas producing anti-HM1.24 antibody were obtained according to the method of Goto, T. et al. (Blood (1994) 84, 1922-1930).

To a BALB/c mouse (manufactured by CLEA Japan) that

previously received intraperitoneal administration of 500  $\mu$ l each of 2,6,10,14-tetramethyl pentadecane (manufactured by Wako Pure Chemical Industries, Ltd.) 11 and 3 days before,  $5 \times 10^6$  hybridoma cells were intraperitoneally injected. From day 10 after the injection of hybridoma cells, the ascites that accumulated in the abdominal cavity of the mouse was collected via a 19-gauge indwelling needle Happycas (manufactured by Medikit). The collected ascites was centrifuged twice at a revolving speed of 1000 and 3000 rpm using a low-speed centrifuge RLX-131 (manufactured by Tomy Seiko) to remove the hybridoma, contaminants such as blood cells and the like.

2. Purification of anti-HM1.24 antibody from mouse ascites

Purification of anti-HM1.24 antibody from the above mouse ascites was conducted in the following method. After adding an equal amount of PBS(-) to the mouse ascites, the mixture was filtered using a hollow fiber filter Mediaprep (manufactured by MILLIPORE) and then was affinity purified using a high speed antibody purification instrument ConSep LC100 (manufactured by MILLIPORE) and the Hyper D Protein A column (column volume 20 ml, manufactured by Nihon Gaisi), and PBS(-) as the adsorption buffer and 0.1 M sodium citrate buffer (pH 4) as the elution buffer according to the attached instructions. The eluted fractions were immediately adjusted to about pH 7.4 by adding 1 M Tris-HCl (pH 8.0), and then were subjected to concentration and buffer replacement to PBS(-) using a centrifuge ultrafiltration concentrator Centriprep 10, which was then filter-sterilized with a membrane filter MILLEX-GV (manufactured by MILLIPORE) having a pore size of 0.22  $\mu$ m to obtain the purified anti-HM1.24 antibody.

3. Purification of control mouse IgG2a

Control mouse IgG2a was purified in the following

method. Commercially available IgG2a (KAPPA) (UPC 10) ascites (manufactured by CAPPEL) was dissolved in purified water and PBS(-). The solution was filtered using a membrane filter Acrodisc (manufactured by Gelman) having a pore size of 0.2  $\mu$ m, and then was affinity-purified using a high speed antibody purification instrument ConSep LC100 (manufactured by MILLIPORE) and the Hyper D Protein A column (column volume 20 ml, manufactured by Nihon Gaisi), and PBS(-) as the adsorption buffer and 0.1 M sodium citrate buffer (pH 4) as the elution buffer according to the attached instructions.

The eluted fractions were immediately adjusted to about pH 7.4 by adding 1 M Tris-HCl (pH 8.0), and then were subjected to concentration and buffer replacement to PBS(-) using a centrifuge ultrafiltration concentrator Centriprep 10, which was then filter-sterilized with a membrane filter MILLEX-GV (manufactured by MILLIPORE) having a pore size of 0.22  $\mu$ m to obtain the purified control mouse IgG2a.

#### 4. Determination of antibody concentration

The concentration of the purified antibody was determined by the measurement of absorbance. Thus, the purified antibody was diluted in PBS(-), the absorbance at 280 nm was measured, and the concentration was calculated using 1.35 OD at 1 mg/ml.

#### Example 2. Effects of anti-HM1.24 antibody on antibody production by human peripheral blood B cells stimulated by SAC

##### 1. Preparation of the human peripheral blood lymphocyte fraction

Peripheral blood, collected from healthy human donors and diluted 1/2 in PBS(-), was layered onto Ficoll-paque (manufactured by Pharmacia) in a 50 ml centrifuge tube (manufactured by Becton Dickinson). After centrifuge at 450 x g for 40 minutes at room

temperature, the mononuclear cell fraction in the interface was isolated. After the fraction was prepared at a suitable density in a RPMI1640 medium (manufactured by GIBCO) containing 10% fetal bovine serum (manufactured by Moredgate), it was incubated under the condition of 37°C and 5% CO<sub>2</sub> for 1 hour in a plastic petri dish. The procedure was repeated twice to remove the cells attached to the dish. The remaining non-adhering cells were used in the following experiments as the human peripheral blood lymphocyte fraction.

## 2. Activation of human peripheral blood B cells by SAC

B cells in the peripheral blood lymphocytes were activated by incubating the human peripheral blood lymphocytes prepared as above at a density of  $5 \times 10^6$  cells/ml with 0.01% SAC (Pansorbin cells, manufactured by Calbiochem.) in the presence or the absence of 1 ng/ml of IL-6 in a polypropylene tube under the condition of 37°C and 5% CO<sub>2</sub> for 2 days. The SAC-treated lymphocytes were suspended in a RPMI1640 medium containing 10% fetal bovine serum (manufactured by Moredgate), and the suspension at a cell density of  $1 \times 10^5$  cells/200 µl/well was cultured with 20 µg/ml anti-HM1.24 antibody or a control mouse IgG2a in a 96-well culture plate (manufactured by Becton Dickinson) under the condition of 37°C and 5% CO<sub>2</sub> for 6 days, followed by collection of the culture supernatant.

## 3. Quantitation of human IgG

The concentration of IgG in the culture supernatant was measured by a human IgG-specific ELISA. Thus, 100 µl of goat anti-human IgG (manufactured by TAGO) diluted to 1 µg/ml in 0.1 M bicarbonate buffer (pH 9.6) was added to a 96-well immunoplate (manufactured by Nunc), and then incubated at 4 °C overnight to immobilize the antibody. After blocking, 100 µl of appropriately diluted culture

supernatant or human IgG (manufactured by CAPPEL) as a standard was added thereto, and then incubated at room temperature for 1 hour.

After washing the plate, 100  $\mu$ l of 2,000-fold  
5 diluted alkaline phosphatase-labeled anti-human IgG  
(manufactured by CAPPEL) was added to the plate, and then  
incubated at room temperature for 1 hour. After washing  
the plate and adding the substrate solution thereto, it  
was incubated. Absorbance at 405 nm was determined using  
10 the MICROPLATE READER Model 3550 (manufactured by Bio-  
Rad).

4. Effects of anti-HM1.24 antibody on antibody  
production by human peripheral blood B cells  
stimulated by SAC

15 It was revealed, as shown in Fig. 1, that  
stimulation by SAC resulted in enhanced IgG production  
and that the addition of control mouse IgG2a thereto did  
not cause any changes. However, the addition of 20  $\mu$ g/ml  
of anti-HM1.24 antibody completely inhibited IgG  
20 production. It was indicated, therefore, that anti-  
HM1.24 antibody inhibited the activation of B cells.

Example 3. Effects of anti-HM1.24 antibody on blast  
formation by PHA-stimulated human T cells

1. Preparation of human peripheral blood T cells

25 Human peripheral T cells were purified from human  
peripheral blood lymphocytes prepared in the above  
example 2 using the Collect Humm T cell Kit (manufactured  
by Biotex) according to the attached procedures.

2. FCM analysis

30 Purified T cells were suspended in a RPMI1640 medium  
containing 2% fetal bovine serum (manufactured by  
Moregale), and the suspension at a cell density of  $1 \times 10^6$   
cells/ml/well was cultured with 0, 1, and 10  $\mu$ g/ml  
PHA (Phytohemagglutinin, manufactured by Sigma) in a 24-  
35 well culture plate under the condition of 37°C and 5% CO<sub>2</sub>  
for 4 days. Some of these after 4 days of culturing were

resuspended in a RPMI1640 medium containing 2% fetal bovine serum (manufactured by Moredale) but containing no PHA, and were cultured for 3 more days. After washing the cells in PBS(-), 100 µl of anti-HM1.24 antibody or control antibody that were diluted to 25 µg/ml in the FACS buffer (containing 2% fetal bovine serum and 0.1% sodium azide) and were incubated on ice for 30 minutes.

After washing with the FACS buffer, 100 µl of 25 µg/ml FITC-labeled goat anti-mouse antibody (GAM, manufactured by Becton Dickinson) was added thereto, which was then incubated on ice for 30 minutes. After washing with the FACS buffer, the cells were suspended in 600 µl of the FACS buffer, and each cell was measured for its fluorescence intensity using the FACScan (manufactured by Becton Dickinson). The result as shown in Fig. 2 revealed that T cell activation by PHA is accompanied by the expression of HM1.24 antigen on the cells. Furthermore, as shown in Fig. 3, it was indicated that the HM1.24 antigen that was once activated and expressed did not disappear in the following culturing.

### 3. Effects of anti-HM1.24 antibody on blast formation by PHA-stimulated human T cells

The purified T cells were suspended in a RPMI1640 medium containing 2% fetal bovine serum (manufactured by Moredale) and then cultured at a cell density of  $1 \times 10^5$  cells/200 µl/well with 1 µg/ml PHA (Phytohemagglutinin, manufactured by Sigma) and anti-HM1.24 antibody or a control mouse IgG2a in a 96-well culture plate under the condition of 37°C and 5% CO<sub>2</sub> for 4 days. <sup>3</sup>H-thymidine (manufactured by Amersham) was then added at 1 µCi/well and the incorporation of radioactivity after 4 hours was measured using a β-counter (manufactured by Pharmacia).

The result as shown in Fig. 4 revealed that blast formation by PHA-stimulated T cells caused an increase in the incorporation of <sup>3</sup>H-thymidine, and that the addition

thereto of control mouse IgG2a at 20 µg/ml caused no changes while that of anti-HM1.24 antibody at 20 µg/ml inhibited the incorporation of <sup>3</sup>H-thymidine. It was hence indicated that anti-HM1.24 antibody inhibits the activation of T cells.

Reference Example 1. Preparation of hybridomas that produce mouse anti-HM1.24 monoclonal antibody

In accordance with the method of Goto, T. et al., Blood (1994) 84, 1992-1930, hybridomas that produce mouse anti-HM1.24 monoclonal antibody were prepared.

A plasma cell line KPC-32 ( $1 \times 10^7$ ) derived from the bone marrow of a patient with human multiple myeloma (Goto, T. et al., Jpn. J. Clin. Hematol. (1991) 32, 1400) was injected twice to the abdominal cavity of a BALB/c mouse (manufactured by Charles River) every six weeks.

Three days prior to sacrificing the animal,  $1.5 \times 10^6$  KPC-32 were injected to the spleen of the mouse in order to further enhance the antibody-producing ability of the mouse (Goto, T. et al., Tokushima J. Exp. Med. (1990) 37, 89). After sacrificing the animal the spleen was extracted and the extracted organ was subjected to cell fusion with the myeloma cell SP2/0 according to the method of Groth, de St. & Schreidegger (Cancer Research (1981) 41, 3465).

By the Cell ELISA (Posner, M.R. et al., J. Immunol. Methods (1982) 48, 23) using KPC-32, the culture supernatant of the hybridoma was screened for antibody.  $5 \times 10^4$  KPC-32 were suspended in 50 ml of PBS and then was aliquoted to a 96-well plate (U-bottomed, Corning, manufactured by Iwaki), which was then air-dried at 37°C overnight. After blocking with PBS containing 1% bovine serum albumin (BSA), the culture supernatant of the hybridoma was added thereto and incubated at 4°C for 2 hours. Then, peroxidase-labeled anti-mouse IgG goat antibody (manufactured by Zymed) was reacted at 4 °C for



1 hour. After washing, o-phenylene diamine solution (manufactured by Sumitomo Bakelite) was reacted at room temperature for 30 minutes.

5 Reaction was stopped by adding 2 N sulfuric acid and the absorbance was measured at 492 nm using the ELISA reader (manufactured by Bio-Rad). In order to remove the hybridoma that produces antibodies against human immunoglobulin, the culture supernatant of the positive hybridoma had previously been adsorbed to human serum and  
10 the reactivity to other cell lines was screened by ELISA. Positive hybridomas were selected, and their reactivity to various cells were investigated by flow cytometry. The last selected hybridoma clone was cloned twice, which was injected to the abdominal cavity of a pristane-  
15 treated BALB/c mice and ascites was obtained therefrom.

Monoclonal antibodies were purified from the ascites of the mouse by ammonium sulfate precipitation and a Protein A affinity chromatography kit (Ampure PA, manufactured by Amersham). The purified antibodies were  
20 labeled with FITC using the Quick Tag FITC binding kit (manufactured by Boehringer Mannheim).

As a result, monoclonal antibodies produced by 30 hybridoma clones reacted with KPC-32 and RPMI 8226. After cloning, the reactivity of the culture supernatant of these hybridomas with other cell lines or peripheral  
25 blood mononuclear cells was investigated.

Of them, 3 clones produced monoclonal antibodies that specifically reacted with the plasma cell. From among the 3 clones, a hybridoma clone that was most  
30 useful for flow cytometry analysis and had a CDC activity to RPMI 8226 was selected and designated as HM1.24. The subclass of the monoclonal antibody produced by this hybridoma was determined by an ELISA using a subclass-specific anti-mouse rabbit antibody (manufactured by  
35 Zymed). Anti-HM1.24 antibody had a subclass of IgG2a  $\kappa$ . The hybridoma HM1.24 that produces anti-HM1.24 antibody was internationally deposited under the provisions of the

Budapest Treaty as FERM BP-5233 on September 14, 1995 with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan.

Reference Example 2. Preparation of humanized anti-HM1.24 antibody

Humanized anti-HM1.24 antibody was obtained in the following method.

From the hybridoma HM1.24 prepared in Reference example 1, total RNA was prepared by the conventional method. From this, cDNA encoding the V region of mouse antibody was synthesized and amplified by a polymerase chain reaction (PCR) method and the 5'-RACE method. A DNA fragment containing the gene encoding a mouse V region was obtained, which was ligated to each plasmid pUC cloning vector and then introduced into competent E. coli cells to obtain an E. coli transformant. The above plasmid was obtained from the transformant. The nucleotide sequence of the cDNA coding region in the plasmid was determined in the conventional method, and the complementarity determining region (CDR) of each V region was determined.

In order to construct a vector expressing chimeric anti-HM1.24 antibody, cDNA encoding a V region of each of L chain and H chain of a mouse anti-HM1.24 antibody was inserted to the HEF vector. Furthermore, in order to construct humanized anti-HM1.24 antibody, a V region CDR of a mouse anti-HM1.24 antibody was grafted to a human antibody by the CDR grafting method. The L chain of human antibody REI was used as the L chain of human antibody, FRs 1 to 3 of the human antibody HG3 was used for the framework regions (FRs) 1 to 3 as the H chain of human antibody, and FR4 of the human antibody JH6 was used for FR4. The amino acid in the FR of the H chain V region was replaced so that the CDR-transplanted antibody could form a suitable antigen-binding site.

In order to express the gene of the L chain and the H chain of the thus constructed humanized anti-HM1.24 antibody in a mammalian cell, each gene was separately introduced into the HEF vector to construct a vector that expresses the L chain or the H chain of the humanized anti-HM1.24 antibody, respectively.

By simultaneously introducing these two expression vectors into the CHO cells, a cell line that produces humanized anti-HM1.24 antibody was established. The antigen binding activity and the binding inhibition activity of humanized anti-HM1.24 antibody obtained by culturing this cell line was investigated by the Cell ELISA using human amnion membrane cell line WISH. The result indicated that the humanized anti-HM1.24 antibody has an antigen binding activity equal to chimeric antibody, and for the binding inhibition activity using a biotinated mouse anti-HM1.24 antibody as well, it had an activity equal to chimeric antibody or mouse antibody.

Incidentally, E. coli having the plasmid that contains the DNA encoding the L chain V region or the H chain V region of chimeric anti-HM1.24 antibody has been internationally deposited under the provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pUC19-1.24L-gk) and Escherichia coli DH5 $\alpha$  (pUC19-1.24H-gyl) on August 29, 1996 with the National Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-5646 and FERM BP-5644, respectively.

Furthermore, E. coli having the plasmid that contains the DNA encoding the version a (SEQ ID NO: 2) of the L chain V region or the version r (SEQ ID NO: 3) of the H chain V region of humanized anti-HM1.24 antibody has been internationally deposited under the provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pUC19-RVLa-AHM-gk) and Escherichia coli DH5 $\alpha$  (pUC19-RVHr-AHM-gyl), respectively, on August 29, 1996 with the National

Institute of Bioscience and Human-Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-5645 and FERM BP-5643, respectively.

5 Furthermore, E. coli having the plasmid that contains the DNA encoding the version s (SEQ ID NO: 4) of the H chain V region of humanized anti-HM1.24 antibody has been internationally deposited under the provisions of the Budapest Treaty as Escherichia coli DH5 $\alpha$  (pUC19-  
10 RVHS-AHM-gyl) on September 29, 1997 with the National Institute of Bioscience and Human Technology, Agency of Industrial Science and Technology, of 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan, as FERM BP-6127.

15 Reference Example 3. Cloning of cDNA encoding HM1.24 antigen protein

cDNA encoding HM1.24 antigen protein specifically recognized by anti-HM1.24 antibody was cloned.

1. Construction of cDNA library

20 1) Preparation of total RNA

From the human multiple myeloma cell line KPMM2, total RNA was prepared according to the method of Chirgwin et al. (Biochemistry, 18, 5294 (1970)). Thus,  $2.2 \times 10^8$  KPMM2 was completely homogenized in 20 ml of 4  
25 M guanidine thiocyanate (manufactured by Nacalai Tesque Inc.). The homogenate was layered on a 5.3 M cesium chloride solution in a centrifuge tube, which was then centrifuged in a Beckman SW40 rotor at 31,000 rpm at 20 °C for 24 hours to precipitate RNA.

30 The RNA precipitate was washed in 70% ethanol and then dissolved in 300  $\mu$ l of 10 mM Tris-HCl (pH 7.4) containing 1 mM EDTA and 0.5% SDS. Pronase (manufactured by Boehringer) was added thereto to a concentration of 0.5 mg/ml and then was incubated at 37°C for 30 minutes.  
35 The mixture was extracted with phenol and chloroform, and RNA was precipitated with ethanol. The RNA precipitate

was then dissolved in 200  $\mu$ l of 10 mM Tris-HCl (pH 7.4) containing 1 mM EDTA.

## 2) Preparation of poly(A)+RNA

5 Poly(A)+RNA was purified using as material 500  $\mu$ g of the total RNA prepared as described above by the Fast Track 2.0 mRNA Isolation Kit (manufactured by Invitrogen) according to the regimen attached to the kit.

## 3) Construction of cDNA library

10 Double stranded cDNA was synthesized using, as material, 10  $\mu$ g of the above poly(A)+RNA prepared by the cDNA synthesis kit TimeSaver cDNA Synthesis Kit (manufactured by Pharmacia) according to the regimen attached to the kit, and was further ligated to the EcoRI adapter supplied in the kit using the Directional Cloning  
15 Toolbox (manufactured by Pharmacia) according to the regimen attached to the kit. The kination and the restriction enzyme NotI treatment of the EcoRI adapter were carried out according to the regimen attached to the kit. Furthermore, the adapter-added double stranded cDNA  
20 having a size of about 500 bp or greater was separated and purified using a 1.5% low boiling point agarose gel (manufactured by Sigma) to obtain about 40  $\mu$ l of adapter-added double stranded cDNA.

The adapter-added double stranded cDNA thus  
25 constructed was ligated using pCOS1 vector (Japanese Patent Application (Kokai) 8-255196) and T4 DNA ligase (manufactured by GIBCO-BRL) that had previously been treated with restriction enzymes EcoRI and NotI and alkaline phosphatase (manufactured by Takara Shuzo) to  
30 construct a cDNA library. The constructed cDNA library was transduced to an E. coli strain DH5 $\alpha$  (manufactured by GIBCO-BRL) and consequently it was estimated to be an independent clone having a total size of about  $2.5 \times 10^6$ .

## 2. Cloning by the direct expression method

35 1) Transfection to COS-7 cells

About  $5 \times 10^5$  clones of the above transduced E. coli were cultured in a 2-YT medium (Molecular Cloning: A Laboratory Manual, Sambrook et al., Cold Spring Harbor Laboratory Press (1989)) containing 50 µg/ml ampicillin to amplify cDNA, which was subjected to the alkali method (Molecular Cloning: A Laboratory Manual, Sambrook et al., Cold Spring Harbor Laboratory Press (1989)) to recover plasmid DNA from the E. coli. The plasmid DNA thus obtained was transfected to COS-7 cells by the electroporation method using the Gene Pulser instrument (manufactured by Bio-Rad).

Thus, 10 µg of the purified plasmid DNA was added to 0.8 ml of the COS-7 cell solution in which the cells had been suspended in PBS at  $1 \times 10^7$  cells/ml, and the mixture was subjected to pulses of 1500 V and 25 µFD capacity. After 10 minutes of a recovery period at room temperature, the electroporated cells were cultured in a DMEM culture medium (manufactured by GIBCO-BRL) containing 10% fetal bovine serum (manufactured by GIBCO-BRL) under the condition of 37°C and 5% CO<sub>2</sub> for 3 days.

## 2) Preparation of a panning dish

A panning dish on which mouse anti-HM1.24 antibody were coated was prepared by the method of B. Seed et al. (Proc. Natl. Acad. Sci. U.S.A., 84, 3365-3369 (1987)). Thus, mouse anti-HM1.24 antibody was added to 50 mM Tris-HCl (pH 9.5) to a concentration of 10 µg/ml. Three milliliters of the antibody solution thus prepared was added to a cell culture dish with a diameter of 60 mm and was incubated at room temperature for 2 hours. After washing three times in 0.15 M NaCl solution, PBS containing 5% fetal bovine serum, 1 mM EDTA, and 0.02% NaN<sub>3</sub> was added to the dish. After blocking, it was used for the following cloning.

## 3) Cloning of cDNA library

The COS-7 cells transfected as described above were peeled off with PBS containing 5 mM EDTA. After washing

the cells once in PBS containing 5% fetal bovine serum, they were suspended in PBS containing 5% fetal bovine serum and 0.02% NaN<sub>3</sub> to a concentration of about 1 x 10<sup>6</sup> cells/ml. The suspension was added to the panning dish prepared as described above and was incubated at room temperature for 2 hours. After gently washing three times in PBS containing 5% fetal bovine serum and 0.02% NaN<sub>3</sub>, plasmid DNA was recovered from the cells bound to the panning dish using a solution containing 0.6% SDS and 10 mM EDTA.

The recovered plasmid DNA was transduced into E. coli DH5α. After amplifying as described above, the plasmid DNA was recovered by the alkali method. The recovered plasmid DNA was transfected into COS-7 cells by the electroporation method and plasmid DNA recovered from the cells bound as described above. A similar procedure was repeated once, and the recovered plasmid DNA was digested with restriction enzymes EcoRI and NotI, thereby confirming the concentration of an insert having a size of about 0.9 kbp.

Furthermore, E. coli cells in which a portion of the recovered plasmid DNA had been transduced were inoculated to a 2-YT agar plate containing 50 µg/ml of ampicillin. After culturing overnight, plasmid DNA was recovered from a single colony. It was digested with restriction enzymes EcoRI and NotI to obtain a clone p3.19 in which the size of the insert is about 0.9 kbp.

This clone was reacted using the PRISM, Dye Terminator Cycle Sequencing kit (manufactured by Perkin Elmer) according to the regimen attached to the kit, and the base sequence was determined using the ABI 373A DNA Sequencer (manufactured by Perkin Elmer). This base sequence and the corresponding amino acid sequence are shown in SEQ ID NO: 1.

#### Industrial Applicability

It was observed that the treatment of anti-HM1.24

antibody caused the inhibition of antibody production by B cells and the inhibition of blast formation by T cells. These facts indicate that anti-HM1.24 antibody has the effect of inhibiting the activation of lymphocytes.

5           Reference to the microorganisms deposited under the Patent Cooperation Treaty, Rule 13-2, and the name of the Depository Institute

          Name: the National Institute of Bioscience and Human  
10           Technology, Agency of Industrial Science and Technology

          Address: 1-3, Higashi 1-chome, Tsukuba city, Ibaraki pref., Japan

Microorganism (1)

15           Name: Escherichia coli DH5 $\alpha$  (pRS38-pUC19)

          Accession number: FERM BP-4434

          Date deposited: October 5, 1993

Microorganism (2)

          Name: hybridoma HM1.24

20           Accession number: FERM BP-5233

          Date deposited: September 14, 1995

Microorganism (3)

          Name: Escherichia coli DH5 $\alpha$  (pUC19-RVHr-AHM-gyl)

          Accession number: FERM BP-5643

25           Date deposited: August 29, 1996

Microorganism (4)

          Name: Escherichia coli DH5 $\alpha$  (pUC19-1.24H-gyl)

          Accession number: FERM BP-5644

          Date deposited: August 29, 1996

30           Microorganism (5)

          Name: Escherichia coli DH5 $\alpha$  (pUC19-RVLa-AHM-gk)

          Accession number: FERM BP-5645

          Date deposited: August 29, 1996

Microorganism (6)

35           Name: Escherichia coli DH5 $\alpha$  (pUC19-1.24L-gk)



Accession number: FERM BP-5646

Date deposited: August 29, 1996

Microorganism (7)

Name: *Escherichia coli* DH5α (pUC19-RVHs-AHM-gyl)

5

Accession number: FERM BP-6127

Date deposited: September 29, 1997